atom concerned has the  $\alpha$ -configuration. In an  $\alpha$ -D-fructopyranose the 2-OH is in the  $\alpha$ -orientation and the 2-CH<sub>2</sub>OH in the  $\beta$ -orientation. The carbon centers in  $\alpha$ -D-glucose, as shown in formula I, are:  $1\alpha$ ,  $2\alpha$ ,  $3\beta$ ,  $4\alpha$ ,  $5\beta$ .

In the Fischer-Rosanoff system of nomenclature as defined by Hudson,<sup>6</sup> the enantiomer of  $\alpha$ -D-glucose of formula II is called  $\alpha$ -L-glucose, but the true meaning is conveyed more accurately by the designation " $\alpha$ "-L-glucose, for the prefix is used merely in a trivial sense and does not bear the connotation of configuration. Since the actual configurations are established with certainty, it would seem rational to abandon the trivial designations in favor of prefixes that have configurational significance. The enantiomer of  $\alpha$ -D-glucose is the exact optical opposite at all centers, including C<sub>1</sub>, and is properly described as:  $1\beta$ ,  $2\beta$ ,  $3\alpha$ ,  $4\beta$ ,  $5\alpha$ .

The proposed system can be distinguished from that in current use by including in the name a number indicating the carbon atom whose configuration is defined by  $\alpha$  or  $\beta$ . Thus I is  $1\alpha$ -Dglucose (or  $1\alpha$ -D-glucopyranose); II is  $1\beta$ -L-glucose; methyl  $\alpha$ -D-fructopyranoside is more specifically described as methyl  $2\alpha$ -D-fructopyranoside. For purposes of discussion it may be convenient to state that  $1\alpha$ -D-mannopyranose is the  $2\beta$ -epimer of  $1\alpha$ -D-glucopyranose, or that methyl  $1\alpha$ -L-idopyranoside is the  $5\alpha$ -epimer of methyl  $1\alpha$ -D-glucopyra-The important relationship discovered by noside. Hudson<sup>1</sup> can be so stated that one rule is applicable to all types of cyclic sugars of both the D- and Lseries: in a pair of glycosidic epimers, the isomer with the glycosidic hydroxyl or alkoxyl group in the  $\alpha$ -orientation is invariably more dextrorotatory than the  $\beta$ -epimer. A relationship discovered by Isbell<sup>7</sup> can be stated as follows: a 1 $\alpha$ -D-pyranose, in which C<sub>1</sub> and C<sub>5</sub> are  $\alpha$  and  $\beta$ , respectively, or *trans*, is oxidized much more slowly than the  $1\beta$ -epimer in which they are  $\beta$ and  $\beta$ , or cis.

(6) C. S. Hudson, "Historical Aspects of Emil Fischer's Fundamental Conventions for Writing Stereo-Formulas in a Plane," *Adv. in Carbohydrate Chem.*, **3**, 1 (1948).

(7) H. S. Isbell, J. Research Natl. Bur. Standards, 18, 505 (1937), RP 990; H. S. Isbell and W. W. Pigman, *ibid.*, 18, 141 (1937) RP 969; H. S. Isbell, J. Chem. Educ., 12, 96 (1935).

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## Polytetramethylene Sebacate: Pyrophoric Lead as an Ester Interchange Catalyst

By C. S. MARVEL\* AND JOHN H. JOHNSON<sup>1</sup>

In preparing polytetramethylene sebacate from butane-1,4-diol and dimethyl sebacate with a litharge catalyst, one run gave a better grade of polyester than had been obtained before. In this polyester sample there was a black deposit that

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(1) Allied Chemical and Dye Corporation Fellow, 1948-1949.

appeared to be finely divided lead. As a result, pyrophoric lead<sup>2</sup> was tried as a catalyst in this interchange reaction and found to work well. The use of a wide variety of finely divided metals as catalysts for the preparation of polyethylene terephthalate from ethylene glycol and dimethyl terephthalate has been described<sup>8</sup> but no mention was made of pyrophoric lead.

The polyesters were prepared in an apparatus similar to that described by Hardy<sup>4</sup> from 11.8 g. of dimethyl sebacate and 5.0 g. of butane-1,4-diol. The charges were heated with the 0.1 g. of catalyst under varying conditions and then the polyesters were purified by solution in chloroform, filtration and reprecipitation with acetone. The following cases seem sufficient to indicate that pyrophoric lead is a satisfactory catalyst in this type of reaction. The yields are essentially quantitative.

**Run No. 1:** 0.1 g. of litharge as catalyst; reaction mixture heated for two hours at 183° and atmospheric pressure, then for one hour at 259° and 0.1 mm. pressure. The polymer was isolated and the intrinsic viscosity taken in 0.4% solution in chloroform at 25.5°; [n] 0.61.

in 0.4% solution in chloroform at 25.5°;  $[\eta]$  0.61. **Run No.** 2: Same catalyst as No. 1; reaction mixture heated three hours at 155° and atmospheric pressure and one hour at 155° and 0.03 mm. pressure;  $[\eta]$  0.33.

**Run No. 3:** 0.1 g. of pyrophoric lead; reaction mixture heated two hours at 172° and atmospheric pressure and six hours longer at 1 mm;  $[\eta]$  0.98.

**Run No. 4**: Same catalyst as No. 3; reaction mixture heated three hours at  $172^{\circ}$  and atmospheric pressure, then three hours at 0.1 mm.;  $[\eta] 0.55$ .

Anal. Caled. for  $C_{14}H_{24}O_4$ : C, 65.60; H, 9.44. Found: C, 65.45; H, 9.63.

The polyesters obtained with pyrophoric lead were whiter than those from the litharge runs. The polymers all melted at  $64-64.5^{\circ}$ .

(2) King, "Inorganic Preparations," D. Van Nostrand Company, New York, N. Y., 1936, p. 24.

(3) British Patent, 578,079.

(4) Hardy, J. Soc. Chem. Ind., 67, 426 (1948).

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## Preparation of Naphthyl Acid Phosphates<sup>1</sup>

By Orrie M. Friedman\* and Arnold M. Seligman<sup>†</sup>

Methods for the histochemical demonstration of alkaline phosphatase<sup>2</sup> and acid phosphatase<sup>3</sup> have been developed utilizing as substrates the calcium salts of  $\alpha$ - and  $\beta$ -naphthyl acid phosphates. Following enzymatic hydrolysis of these substrates, coupling with a suitable diazonium compound, results in the deposition of an insoluble azo dye at the site of enzymatic activity.

Although the calcium salts of these phosphoric esters are readily prepared<sup>2,3</sup> their poor solubility in water requires their use in a fine suspension for

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(1) This investigation was supported by a research grant from the National Cancer Institute of the National Institutes of Health, Public Health Service.

(2) (a) Menten, Junge and Green, J. Biol. Chem., 153, 471 (1944).
(b) Manheimer and Seligman, J. Nat. Concer Instit., 9, 181 (1948).

(3) Seligman and Manheimer, *ibid.*, 9, 427 (1949).